

Effect of salt and pH on the growth and production of alkaline proteases from Haloalkaliphilic bacteria isolated from saline desert

Hitarth B. Bhatt, Pooja M. Palanpura and Satya P. Singh*

UGC-CAS Department of Biosciences, Saurashtra University, Rajkot-360005

*Corresponding Authors: Prof. Satya P. Singh, E-mail: satyapsingh@yahoo.com

ABSTRACT

Saline desert of Kutch is yet unexplored for its microbial diversity and biotechnological potential. Proteases are significant with respect to their applications and cellular roles. A total of 15 bacteria isolated from the Little Rann of Kutch were isolated and screened for their growth and protease secretion at different pH and salt concentrations. Effect of NaCl concentrations and pH was assessed on growth and production of the proteases. Overall, 50 % of the isolates grew in the range of 0-20 % (w/v) NaCl, while nearly 45 % of them optimally produced extracellular proteases at 10 % (w/v) NaCl and 20 % isolates produced protease optimally at 15% (w/v) NaCl concentration. Majority of the isolates secreted protease in pH range 7-11 and nearly 70 % the isolates produced enzyme in the broad range of pH, 7-11 and nearly 6 % isolates produced protease in the range of pH 9-11. Further, around 40% of the isolates produced proteases optimally at pH 9-11. Therefore, the isolates demonstrated a broad range of pH and salt for growth and protease production. The study suggests wide prospective of the applications of these organisms and their enzymes.

Keywords: *Haloalkaliphilic bacteria; saline desert; salt tolerance; enzymatic potential; alkaline protease*

INTRODUCTION

Deserts being arid regions receive <250 mm average rains. Therefore, a number of studies have been carried out for microbial diversity and biotechnological potential. Despite the large land area of the arid regions, only limited is known about the microbial communities ^[1, 2].

Further, Indian deserts are least explored for their microbial community and enzymatic potential ^[2, 3]. The saline desert of Kutch is unique as the north head of the Gulf of Kutch adjoins the Desert of Kutch with the regular flow of saline water during tides or through the water drifted by the south-west winds. Rainfall is limited, so that as water recedes and evaporates, there appears a crust of halite and gypsum crystals which grow in the clay and sands ^[4]. Thus, the study of microbes from this unique habitat adds to our understanding on the adaptation, ecology and enzymatic potential.

The microflora of the saline ecosystems has attracted a great deal of attention during the last couple of decades. Halophilic and halotolerant bacteria belong to a class of extremophilic organisms that live in habitats of high ionic strength, such as marine water, salt lakes, brines, saline deserts, salterns, saline soils, and salted foods ^[1, 5-10]. These microbes possess adaptation mechanisms to thrive under the dual extremities of high salinity and alkaline pH ^[11]. From a biotechnological stand point, enzymes withstanding multitude of the extremity from the haloalkaliphilic bacteria can be quite useful in various applications ^[6, 12-15].

To explore newer habitats for novel metabolites and bioactive molecules, there is a constant need to explore unexplored or unusual habitats. The arid or semiarid regions hold significance as they harbour an array of unique organisms with unexplored metabolic potential. In this study, we investigated the effect of salt and pH on the growth and protease production in the bacteria isolated from the saline desert of Kutch.

MATERIALS AND METHODS

Isolates of the organisms

Four different media SP medium consisting, (g/l): (NaCl, 49; KCl, 1; MgSO₄.7H₂O, 0.5; CaCl₂.2H₂O, 0.18; NaHCO₃, 0.03; NaBr, 0.115; FeCl₃.6H₂O, 0.5; Bacto trypton, 5; Yeast extract, 10; Glucose, 1; Agar, 3%), Reasoner's 2A (R2A) agar (HiMedia Laboratories, India) consisting, (g/l): (Casein acid hydrolysate, 0.5; yeast extract, 0.5; protease peptone, 0.5; dextrose, 0.5; starch, 0.5; KH₂PO₄, 0.3; MgSO₄, 0.024; Sodium pyruvate, 0.30; NaCl (5%, w/v)), Soil Extract medium (SE) consisting , (g/l): (Glucose,1; K₂HPO₄, 0.5; Soil extract, 17.75, Agar, 15) and HM medium consisting, (g/l): NaCl, 220; MgSO₄.7H₂O, 10; KCl, 5; Sodium citrate, 3; KNO₃, 1; CaCl₂.2H₂O, 0.20, Trace minerals, 0.5; Bacto tryptone, 5; Yeast extract, 1; Agar, 3% (w/v) were used to target and isolate diverse microbiota of desert soil.

Total of 15 isolates were obtained from Venasar study site which were further selected for the optimization of protease secretion. Media profile of the isolates is given in Table 1.

Effect of salt on the growth and protease production

Actively growing cultures of 15 bacteria isolates, prepared in their respective isolation medium at pH 8 were used as inoculums for the assessment of the production of the alkaline protease. The regular spots of cultures were inoculated on the gelatin agar medium containing (g/l): gelatin, 30; peptone, 10; agar, 30; NaCl (0%, 5%, 10%, 15% and 20% (w/v)) and pH was kept 8 by adding separately autoclaved Na₂CO₃ in the medium. The inoculated plates were incubated for 72-96 hours at 37°C. Frazier's reagent (g/l: HgCl₂, 150gm; Concentrated HCl, 200mL) was poured into the plate for the detection of protease production. The extracellular protease secretion was indicated by the clear zone of substrate utilization surrounding the colonies. The colony diameter and zone of clearance were measured. The ratio of zone diameter to the colony diameter was calculated to estimate the relative enzyme production as a function of the colony size.

Effect of pH on the growth and protease production

Inoculums of the isolates were prepared as described above. The regular spots of cultures were inoculated on the gelatin agar medium containing (g/l): gelatin, 30; peptone, 10; agar, 30; NaCl (5% and 25% (w/v)) with different pH at interval of 1 (7, 8, 9, 10 and 11). The pH 7, 8 and 9 was adjusted with separately autoclaved Na₂CO₃, while pH 10 and 11 was adjusted by autoclaved 1M NaOH. The plates were incubated for 72-96 hours at 37°C. Detection of protease production was followed as described in above section.

RESULTS

Revival of the isolates

Haloalkalophiles were isolated from the samples collected from the saline desert area from Little Rann of Kutch (Gujrat) on the SP agar, Reasoner's 2A (R2A) agar, Soil Extract medium (SE), HM agar.

Isolates	Isolation Media
VS -1	SP
VS -2	SP
VS -3	SP
VS -5	SP
VS -6	SP
VS -9	SP
VS -10	SP
VS -12	SP
VS -16	R2A
VS -19	R2A
VS -21	SE
VS -22	SE
VS -24	SE
VS -32	HM
VS -35	SP

Table 1 Isolation of all the bacterial isolates of the saline desert of Kutch

Effect of salt on the growth and protease production

The effect of NaCl was studied on the growth and protease production in 15 bacterial isolates. Overall, ~ 50% isolates could grow over a wide range of 0-20% (w/v) NaCl, while ~ 40 % of the isolates could grow in range of 0-15 % (w/v) NaCl and ~ 6% were able to grow in the range of 0-10 % (w/v) NaCl. Isolates VS-1, VS-2 and VS-35 had optimum growth at 10 % (w/v) NaCl concentration, while VS-3, VS-5, VS-12, VS-19 and VS-21 have shown growth optima at 5 % (w/v) NaCl concentration. On the other hand, ~ 30% isolates produced enzyme in the broad range of 0-15 % (w/v) NaCl and 20 % produced protease in the range of 0-10 % (w/v) NaCl. Interestingly, 13% isolates produced protease in the range of 0-20 % (w/v) NaCl.

Further, it was observed that 45 % of the isolates secreted protease optimally at 10 % (w/v) NaCl concentration and 30% of them produced protease optimally at 5 % (w/v) NaCl concentration. In addition, ~ 20 % of the isolates produced protease optimally at 15 % (w/v) NaCl concentration. Isolate VS-1 did not produce protease at any of the salt concentration examined. Overall, many isolates have shown broad salt requirement for both, growth and production of the proteases.

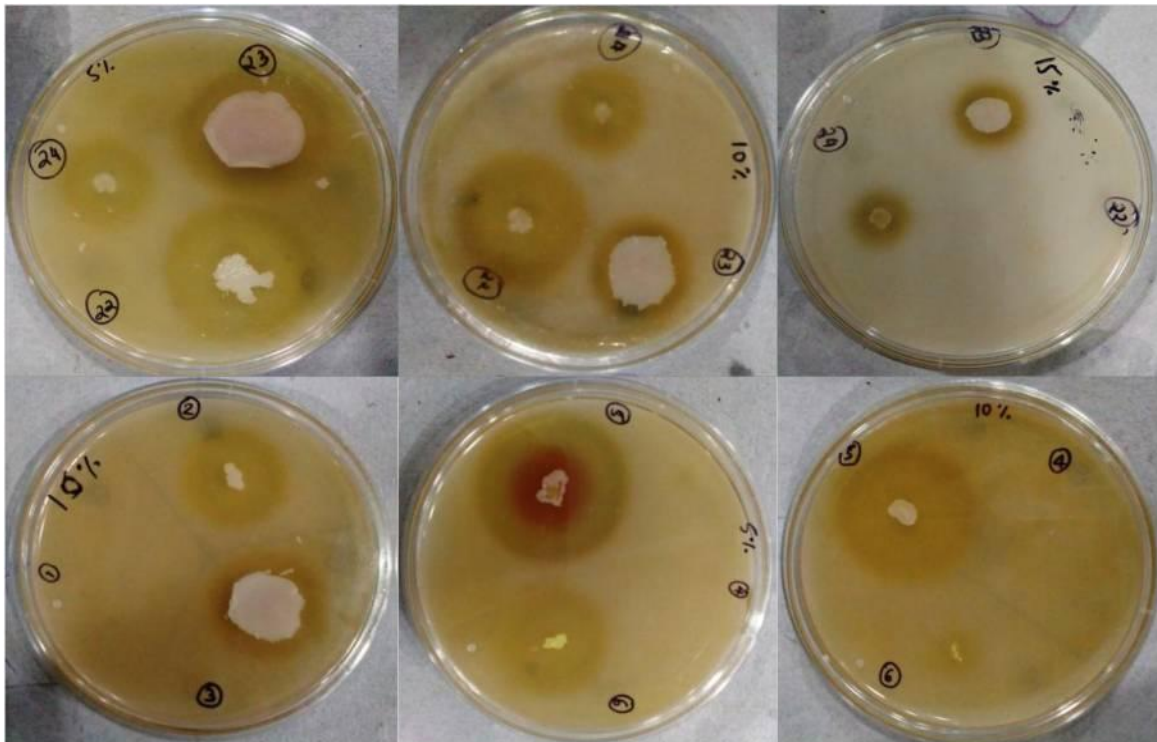


Figure 1 Screening of protease production using a drop spot technique on the gelatine agar plates

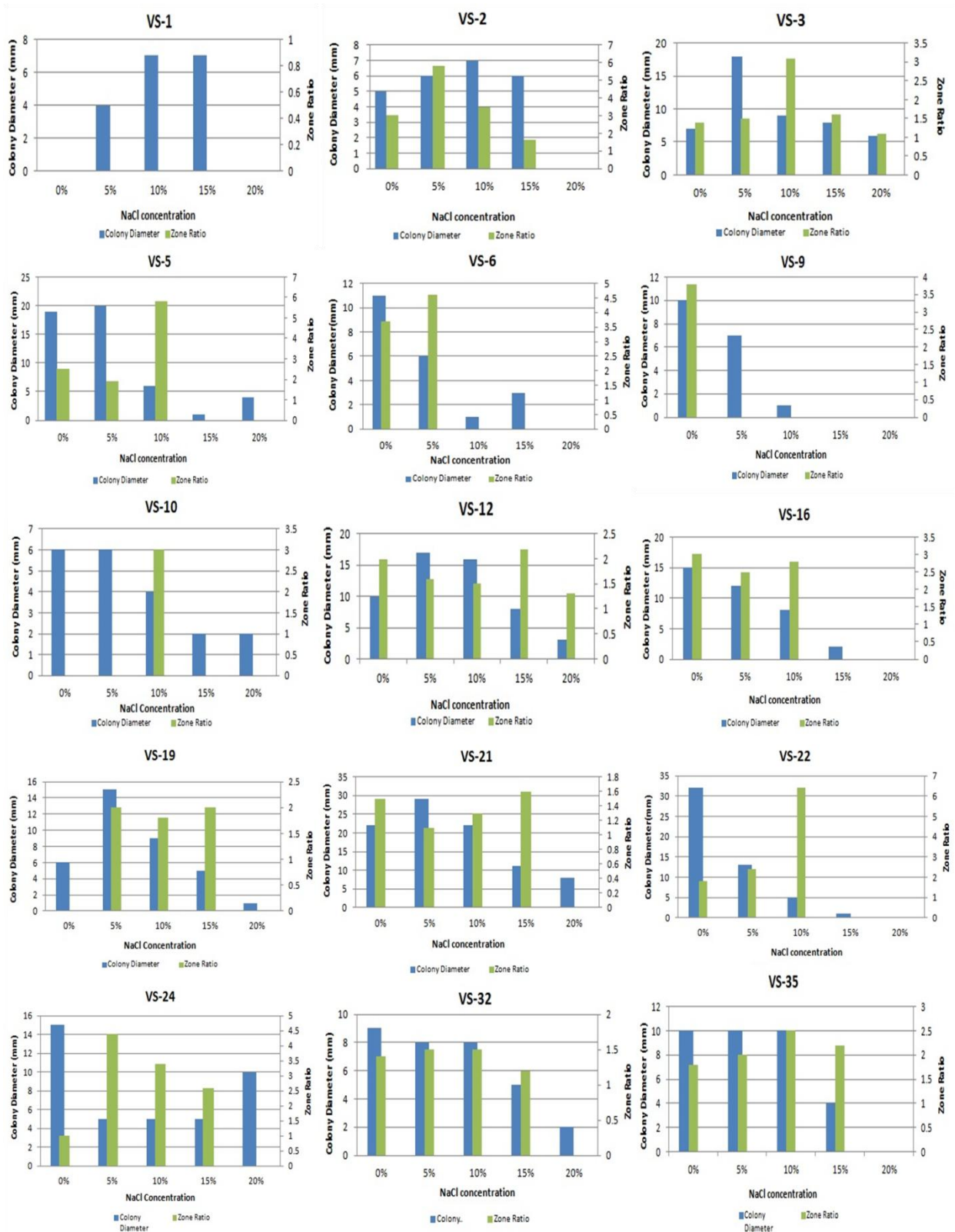


Figure 2 Effect of salt on the growth and production of the proteases from haloalkaliphilic bacteria isolated from saline desert of Kutch

Effect of pH on the growth and production of the alkaline protease

The effect of pH was examined on the growth and protease production of all the 15 isolates. Overall, 90 % of the isolates could grow in the range of pH 7-11, while 6 % of the isolates grew in the range of pH 7-10. Many of the isolates were able to grow optimally in the pH alkaline. Strains VS-2, VS-10, VS-12 and VS-22 have shown growth optima at pH 9 while strains VS-21 and VS-32 had optimum growth at pH 10 and 11, respectively. On the other hand, ~ 70% of the isolates produced enzyme in the broad range of pH 7-11 and 6 % produced proteases in the range of pH 9-11. Further, it was observed that, 40 % the isolates optimally produced protease at pH 11. Further, 40 % of the isolates optimally produced protease at pH 9. In addition, 6 % of the isolates optimally produced protease at pH 7, pH 8 and pH 9 each. Overall, many isolates have shown broad pH requirement not only for their growth but also for the protease secretion.

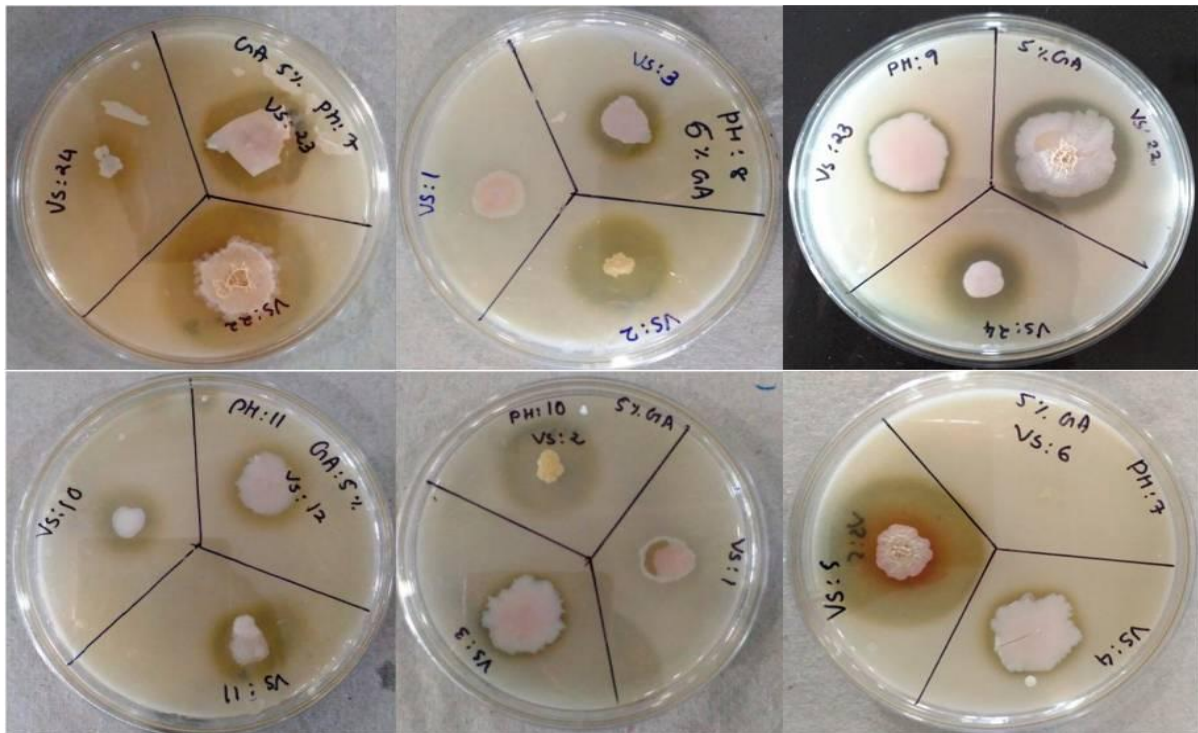


Figure 3 Screening of protease production at different pH using a drop spot technique on gelatine agar plates

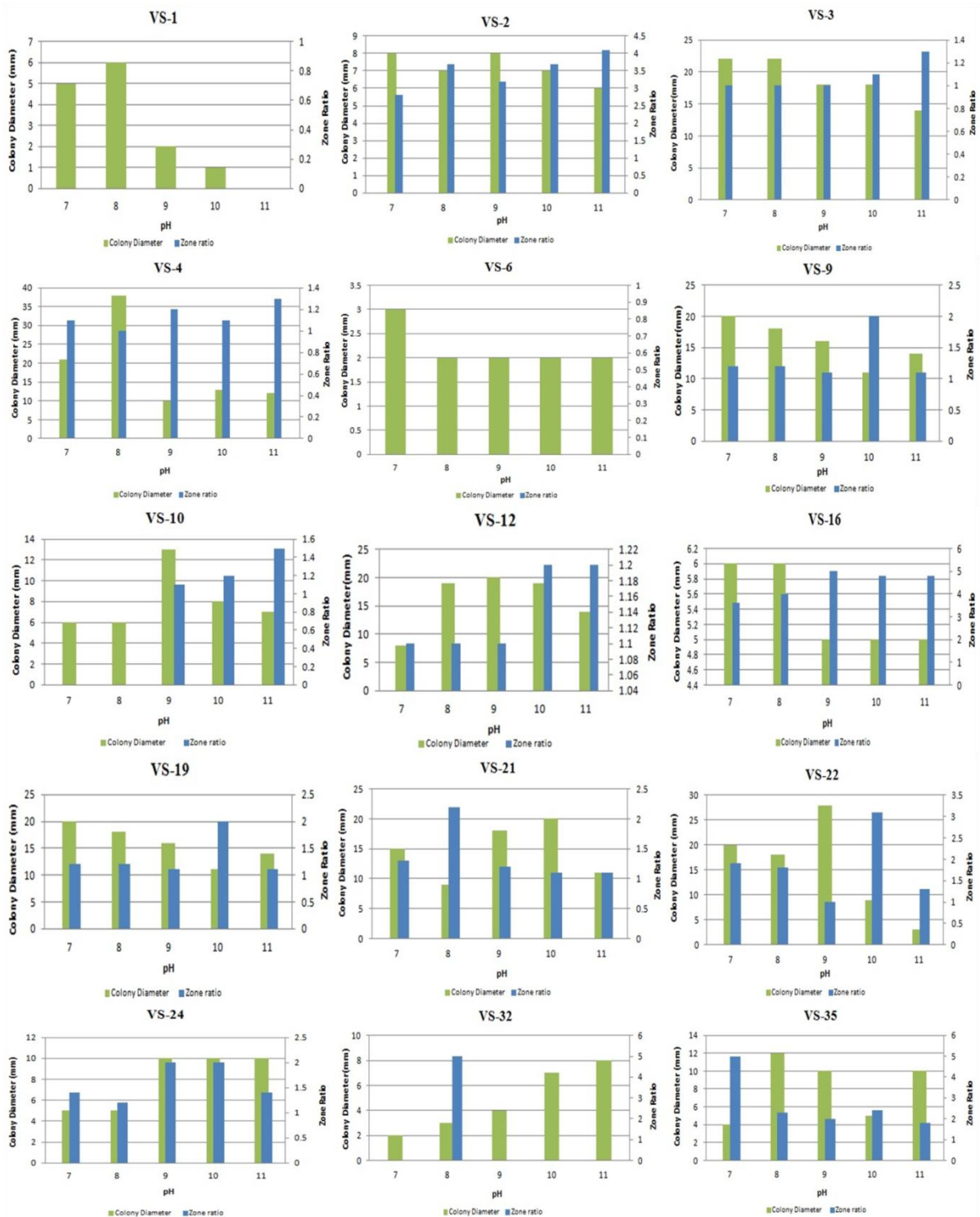


Figure 4 Effect of pH on the growth and production of protease from haloalkaliphilic bacteria isolated from saline desert of Kutch

DISCUSSION

Till now many haloalkaliphilic bacteria and actinomycetes have been studied from various marine environments [15-22]. However, arid environments have been relatively less explored [2,

^{10, 23, 24]}. Further, microbial community analysis have been carried out in deserts such as, Antacama Desert (Chile), Monegros Desert (Spain), Negev desert (Israel), Gobi desert (Mongolia) and Taklamaken desert (China) Sahara desert (Africa), Sonoran desert (America) while Indian desert areas are not much explored for microbial diversity and their biotechnological application point of view ^[1, 2].

In view of these facts, we isolated haloalkaliphilic and haloalkalitolerant bacteria from the desert of Kutch, an unique and unexplored arid zone. It has a special and different demography from other deserts of the world because of its location near the sea and low-lying areas, through which marine water enters into the vast area. Consequently, the desert of Kutch is an admixture of the saline, marshy and coastal desert where water and soils are saline. This characteristic makes desert a special habitat from the biodiversity point of view because it harbours its own, unique environment ^[1, 4]. The study of the microorganisms at the limits of the biological tolerance facilitates to understand the adaptation strategies.

The haloalkaliphilic microorganisms are significant not only due to the necessity for understanding the mechanisms of adaptations, but also due to the huge prospect of their application potential ^[11]. Efforts have focused on the investigations over the past decade on the enzymes capable to function under harsh conditions ^[12, 15, 22, 25, 26]. In present study, Most of the isolates were moderately halophilic and alkaliphilic in nature and hence referred as haloalkaliphiles. Majority of the isolates produced proteases at 0-15% (w/v) NaCl, 10% being optimum in most. Similarly, majority of the isolates produced proteases in broad range of pH, 7-11. This is considered to be an excellent feature as far as protease production flexibility is concerned. Further, many isolates optimally produced proteases at pH 10-11, a range which did not exactly correspond with its growth. So, growth and enzyme production were not linked. By and large, there was considerable difference in the production of the protease among the isolates. Rekik et al. (2019) reported alkaline protease from *Bacillus safensis* active over broad range of pH and salt concentrations ^[27]. Similarly, broad pH and temperature-active protease from *Bacillus subtilis* was reported ^[28].

Enzyme secretion at such a broad pH and salt concentration makes them interesting candidate for varied applications. It is very likely that the proteases produced from these bacteria will be stable at the range of pH and salt concentrations ^[29, 30]. Moreover, dual extremities; alkaline pH and salinity further project them as promising candidates for various biotechnological applications ^[31-34].

CONCLUSIONS

In this report, effect of salt and pH has been assessed on the protease production in the bacteria isolated from the saline desert of Kutch, Gujarat, India. Not many studies are available on the exploration of microbial diversity and biotechnological potential of the desert ecosystems. In this study, broad tolerance to pH and salt was observed for their growth and the growth was not linked with the protease production. Protease production in such a broad pH and salt concentration range increases their prospective potential in varied industrial applications.

ACKNOWLEDGEMENT

HB is thankful to the Council of Scientific and Industrial Research (CSIR), New Delhi, India for the award of Senior Research Fellowship (CSIR–SRF) and UGC, New Delhi for the award of BSR-Meritorious Fellowship. The authors are grateful to the University Grants Commission (UGC), India and Saurashtra University for the infrastructural and financial support. Support under the UGC-Centre of Advanced Study, DST-FIST program and DBT-Multi Institutional Project is also acknowledged.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

1. Bhatt HB, Singh SP. Phylogenetic and phenogram based diversity of haloalkaliphilic bacteria from the saline desert. In *Microbial Biotechnology*. 2017 Mar 16 (pp. 373-386). Apple Academic Press.
2. Bhatt HB, Gohel SD, Singh SP. Phylogeny, novel bacterial lineage and enzymatic potential of haloalkaliphilic bacteria from the saline coastal desert of Little Rann of Kutch, Gujarat, India. *3 Biotech*. 2018 Jan 1;8(1):53.
3. Pandit AS, Joshi MN, Bhargava P, Shaikh I, Ayachit GN, Raj SR, Saxena AK, Bagatharia SB. A snapshot of microbial communities from the Kutch: one of the largest salt deserts in the World. *Extremophiles*. 2015 Sep 1;19(5):973-87.
4. Gupta V, Ansari AA. Geomorphic portrait of the Little Rann of Kutch. *Arabian Journal of Geosciences*. 2014 Feb 1;7(2):527-36.
5. Purohit MK, Singh SP. Comparative analysis of enzymatic stability and amino acid sequences of thermostable alkaline proteases from two haloalkaliphilic bacteria

- isolated from Coastal region of Gujarat, India. International journal of biological macromolecules. 2011 Jul 1;49(1):103-12.
6. Pandey S, Rakholiya KD, Raval VH, Singh SP. Catalysis and stability of an alkaline protease from a haloalkaliphilic bacterium under non-aqueous conditions as a function of pH, salt and temperature. Journal of bioscience and bioengineering. 2012 Sep 1;114(3):251-6.
 7. Raval VH, Purohit MK, Singh SP. Diversity, population dynamics and biocatalytic potential of cultivable and non-cultivable bacterial communities of the saline ecosystems. In Marine Enzymes for Biocatalysis 2013 Jan 1 (pp. 165-189). Woodhead Publishing.
 8. Karray F, Abdallah MB, Kallel N, Hamza M, Fakhfakh M, Sayadi S. Extracellular hydrolytic enzymes produced by halophilic bacteria and archaea isolated from hypersaline lake. Molecular biology reports. 2018 Oct 1;45(5):1297-309.
 9. Boyadzhieva I, Tomova I, Radchenkova N, Kambourova M, Poli A, Vasileva-Tonkova E. Diversity of Heterotrophic Halophilic Bacteria Isolated from Coastal Solar Salterns, Bulgaria and Their Ability to Synthesize Bioactive Molecules with Biotechnological Impact. Microbiology. 2018 Jul 1;87(4):519-28.
 10. Bhatt HB, Begum MA, Chintalapati S, Chintalapati VR, Singh SP. *Desertibacillus haloalkaliphilus* gen. nov., sp. nov., isolated from a saline desert. International journal of systematic and evolutionary microbiology. 2017 Sep 21;67(11):4435-42.
 11. Raval VH, Bhatt HB, Singh SP. Adaptation Strategies in Halophilic Bacteria. In Extremophiles 2018 Jan 9 (pp. 137-164). CRC Press.
 12. Raval VH, Pillai S, Rawal CM, Singh SP. Biochemical and structural characterization of a detergent-stable serine alkaline protease from seawater haloalkaliphilic bacteria. Process Biochemistry. 2014 Jun 1;49(6):955-62.
 13. DasSarma S, DasSarma P. Halophiles and their enzymes: negativity put to good use. Current opinion in microbiology. 2015 Jun 1;25:120-6.
 14. Hammami A, Fakhfakh N, Abdelhedi O, Nasri M, Bayouhdh A. Proteolytic and amylolytic enzymes from a newly isolated *Bacillus mojavensis* SA: characterization and applications as laundry detergent additive and in leather processing. International journal of biological macromolecules. 2018 Mar 1;108:56-68.
 15. Sharma AK, Kikani BA, Singh SP. Biochemical, thermodynamic and structural characteristics of a biotechnologically compatible alkaline protease from a

- haloalkaliphilic, *Nocardiopsis dassonvillei* OK-18. International Journal of Biological Macromolecules. 2020 Mar 5.
16. Peng J, Zhang X, Xu X, He F, Qi S. Diversity and chemical defense role of culturable non-actinobacterial bacteria isolated from the South China Sea gorgonians. J Microbiol Biotechnol. 2013 Apr 1;23(4):437-43.
 17. Gohel SD, Singh SP. Molecular Phylogeny and Diversity of the Salt-Tolerant Alkaliphilic Actinobacteria Inhabiting Coastal Gujarat, India. Geomicrobiology Journal. 2018 Oct 21;35(9):775-89.
 18. Bachran M, Kluge S, Lopez-Fernandez M, Cherkouk A. Microbial diversity in an arid, naturally saline environment. Microbial ecology. 2019 Aug 15;78(2):494-505.
 19. Rathore DS, Sheikh M, Gohel S, Singh SP. Isolation strategies, abundance and characteristics of the marine actinomycetes of Kachhighadi, Gujarat, India. Journal of the Marine Biological Association of India. 2019 Jan;61(1):72.
 20. Osman JR, Regeard C, Badel C, Fernandes G, DuBow MS. Variation of bacterial biodiversity from saline soils and estuary sediments present near the Mediterranean Sea coast of Camargue (France). Antonie van Leeuwenhoek. 2019 Mar 15;112(3):351-65.
 21. Sheikh M, Rathore DS, Gohel S, Singh SP. Cultivation and characteristics of the Marine Actinobacteria from the Sea water of Alang, Bhavnagar.
 22. Thakrar FJ, Singh SP. Catalytic, thermodynamic and structural properties of an immobilized and highly thermostable alkaline protease from a haloalkaliphilic actinobacteria, *Nocardiopsis alba* TATA-5. Bioresource technology. 2019 Apr 1;278:150-8.
 23. Babavalian H, Amoozegar MA, Zahraei S, Rohban R, Shakeri F, Moghaddam MM. Comparison of bacterial biodiversity and enzyme production in three hypersaline lakes; Urmia, Howz-Soltan and Aran-Bidgol. Indian journal of microbiology. 2014 Dec 1;54(4):444-9.
 24. Ruginescu R, Purcărea C, Dorador C, Lavin P, Cojoc R, Neagu S, Lucaci I, Enache M. Exploring the hydrolytic potential of cultured halophilic bacteria isolated from the Atacama Desert. FEMS microbiology letters. 2019 Sep;366(17):fnz224.
 25. Kikani BA, Singh SP. The stability and thermodynamic parameters of a very thermostable and calcium-independent α -amylase from a newly isolated bacterium, *Anoxybacillus beppuensis* TSSC-1. Process Biochemistry. 2012 Dec 1;47(12):1791-8.

26. Shukla RJ, Singh SP. Structural and catalytic properties of immobilized α -amylase from *Laceyella sacchari* TSI-2. International journal of biological macromolecules. 2016 Apr 1;85:208-16.
27. Rekik H, Jaouadi NZ, Gargouri F, Bejar W, Frikha F, Jmal N, Bejar S, Jaouadi B. Production, purification and biochemical characterization of a novel detergent-stable serine alkaline protease from *Bacillus safensis* strain RH12. International journal of biological macromolecules. 2019 Jan 1;121:1227-39.
28. Uttatree S, Charoenpanich J. Isolation and characterization of a broad pH-and temperature-active, solvent and surfactant stable protease from a new strain of *Bacillus subtilis*. Biocatalysis and Agricultural Biotechnology. 2016 Oct 1;8:32-8.
29. Patel RK, Dodia MS, Joshi RH, Singh SP. Production of extracellular halo-alkaline protease from a newly isolated haloalkaliphilic *Bacillus* sp. isolated from seawater in Western India. World Journal of Microbiology and Biotechnology. 2006 Apr 1;22(4):375-82.
30. Dodia MS, Rawal CM, Bhimani HG, Joshi RH, Khare SK, Singh SP. Purification and stability characteristics of an alkaline serine protease from a newly isolated Haloalkaliphilic bacterium sp. AH-6. Journal of industrial microbiology & biotechnology. 2008 Feb 1;35(2):121-31.
31. Raval VH, Rawal CM, Pandey S, Bhatt HB, Dahima BR, Singh SP. Cloning, heterologous expression and structural characterization of an alkaline serine protease from sea water haloalkaliphilic bacterium. Annals of microbiology. 2015 Mar 1;65(1):371-81.
32. Sinha R, Khare SK. Immobilization of halophilic *Bacillus* sp. EMB9 protease on functionalized silica nanoparticles and application in whey protein hydrolysis. Bioprocess and biosystems engineering. 2015 Apr 1;38(4):739-48.
33. Zhou C, Qin H, Chen X, Zhang Y, Xue Y, Ma Y. A novel alkaline protease from alkaliphilic *Idiomarina* sp. C9-1 with potential application for eco-friendly enzymatic dehairing in the leather industry. Scientific reports. 2018 Nov 7;8(1):1-8.
34. Purohit MK, Raval VH, Singh SP. Haloalkaliphilic bacteria: molecular diversity and biotechnological applications. In Geomicrobiology and Biogeochemistry 2014 (pp. 61-79). Springer, Berlin, Heidelberg.